

Electrocardiogram (ECG) IMPC_ECG_002

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Purpose

To provide a high throughput method to obtain Electrocardiograms in a conscious mouse.

Experimental Design

- **Minimum number of animals** : 5-7M + 5-7F
- **Age at test**: Ideal age = 12 weeks \pm 3 days.
- **Sex**: We would expect the results of this test to show sexual dimorphism

Procedure

1. The lead plates are to be snapped into place onto the top of the pre-amplifier tower. The covering is removed to reveal three gel coated pads surrounded by a sticking plate. The plate will need to be covered with the extra cover in the package.
2. Turn on the combined amplifier and the pre-amplifier tower.
3. Double click the icon ECG acquisition on the acquisition computer.
4. Open the ECG set up file (for default settings).
6. Place mouse on pad, lowering the Red Acrylic Cubby to surround the mouse on 3 sides discouraging escape.
7. Press Start.
8. After the desired acquisition time, (5-10 minutes) stop the reading. There will be one long reading.
9. Save the data.
10. For additional readings create a new session using the same settings as before.
11. When saving sections with good readings, highlight the selected area and then save.

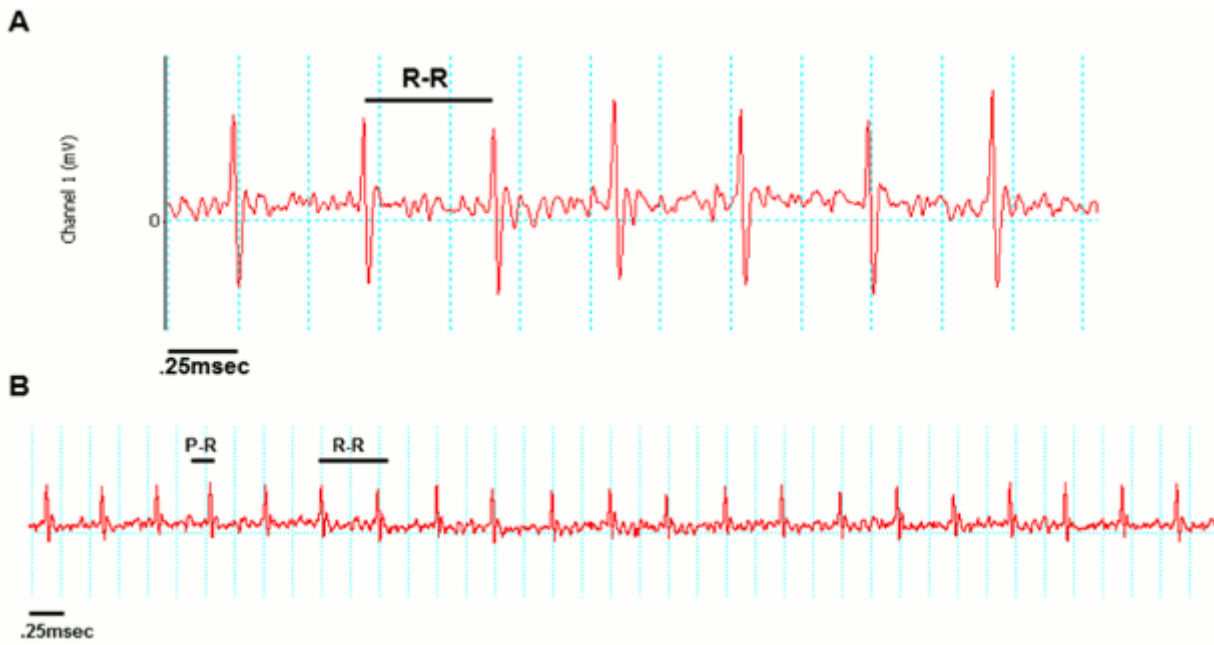
Notes

Data Analysis

1. Open Emouse Analyses icon
2. Select ECG signals
3. Choose folder (all readings in folder will show)
4. Click PNN X (for mice: N-N > than 6 ms)
5. Choose file(s) by highlighting
6. Go
7. Bottom file is the corrected file

8. Red dots should be on peak of R waves, if image appears inverted click invert
9. Click Add, or minus if R waves are not marked with red dots or if too many are marked
 - L click to zoom in
 - R click to zoom out
10. 'What if?' button to remove unwanted sections
 - L click image (zooms in)
 - L click left boundary
 - L click right boundary
11. Options- click more if want to exclude more sections
12. Undo available
13. Go
14. Here can input animal data if desired
15. Save- For the first mouse in in group, hit save, a new results folder will be created within the folder with the mouse data. Then can click quick save or next.
16. For the rest of the mice in the series, can hit quick save at this point- saves in last selected file - will group all files together in same excel sheet.
17. Open Emouse Analyses icon
18. Select ECG signals
19. Choose folder (all readings in folder will show)
20. Click PNN X (for mice: N-N > than 6 ms)
21. Choose file(s) by highlighting
22. Go
23. Bottom file is the corrected file
24. Red dots should be on peak of R waves, if image appears inverted click invert
25. Click Add, or minus if R waves are not marked with red dots or if too many are marked
 - L click to zoom in
 - R click to zoom out
26. 'What if?' button to remove unwanted sections
 - L click image (zooms in)
 - L click left boundary
 - L click right boundary
27. Options- click more if want to exclude more sections
28. Undo available
29. Go
30. Here can input animal data if desired
31. Save- For the first mouse in in group, hit save, a new results folder will be created within the folder with the mouse data. Then can click quick save or next
32. For the rest of the mice in the series, can hit quick save at this point- saves in last selected file - will group all files together in same excel sheet

Examples of good readings



Data QC

Analysis room should be dim and quiet. Keep the door closed preferably while analysis is taking place.

Figure A. Taking a reading

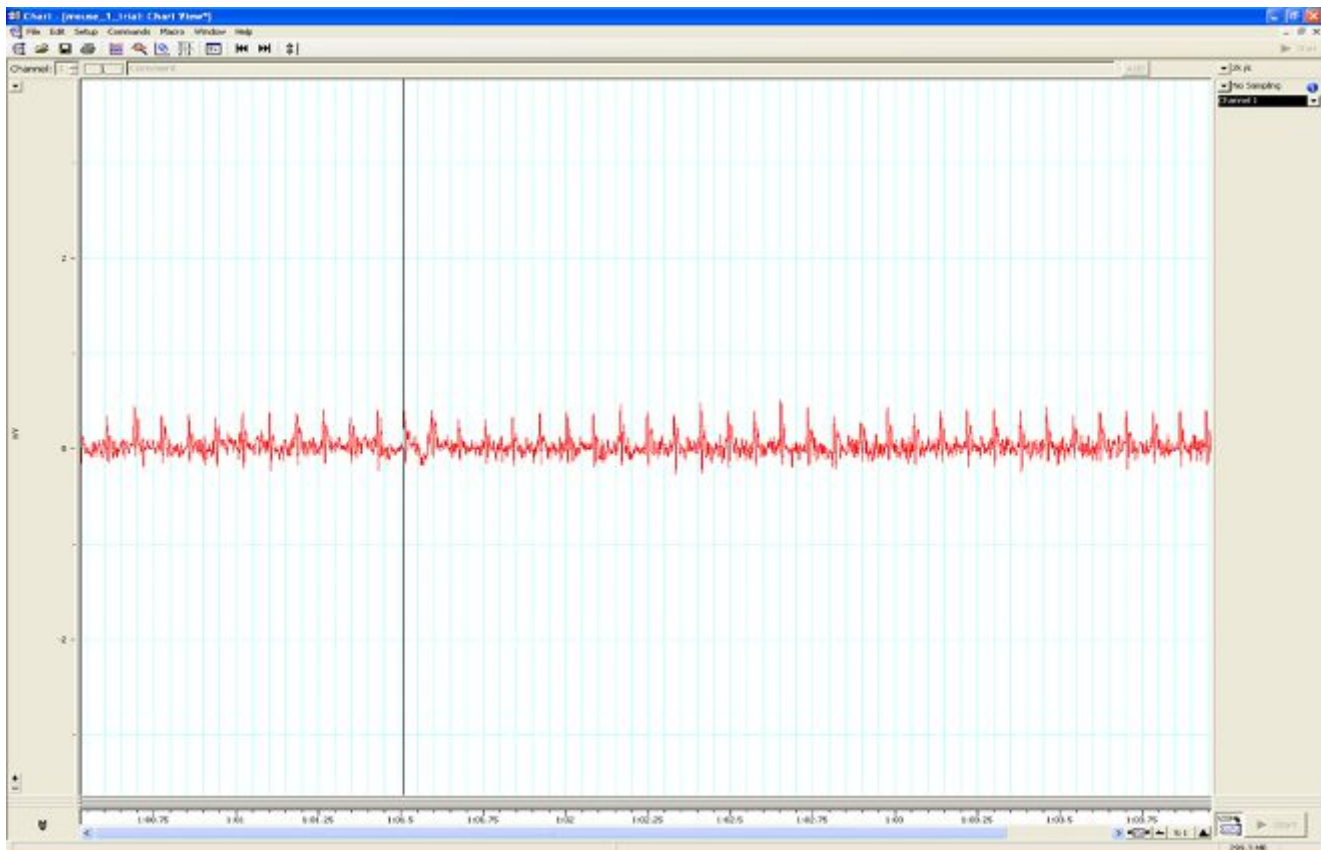


Figure B. Saving a section of the reading

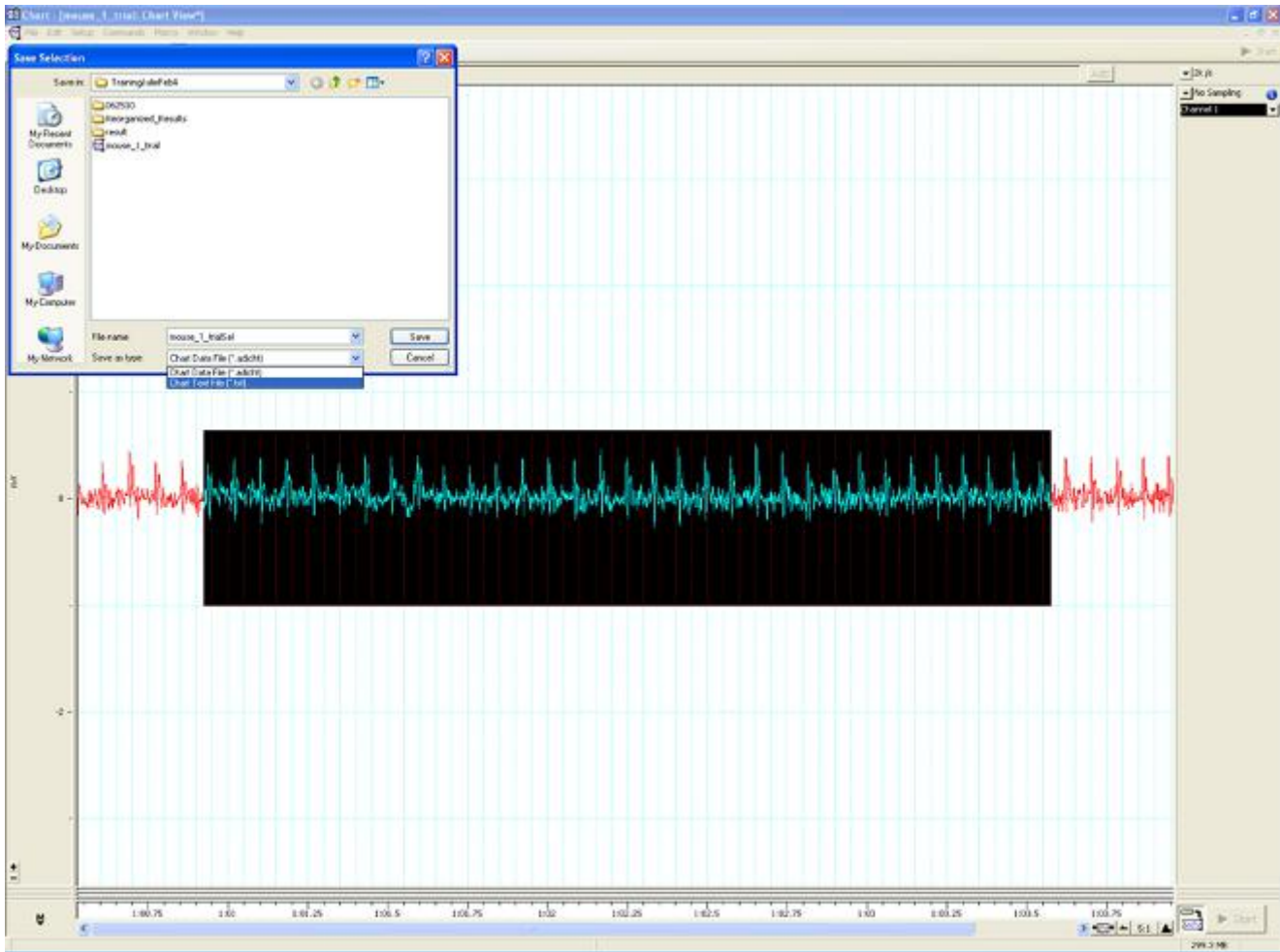
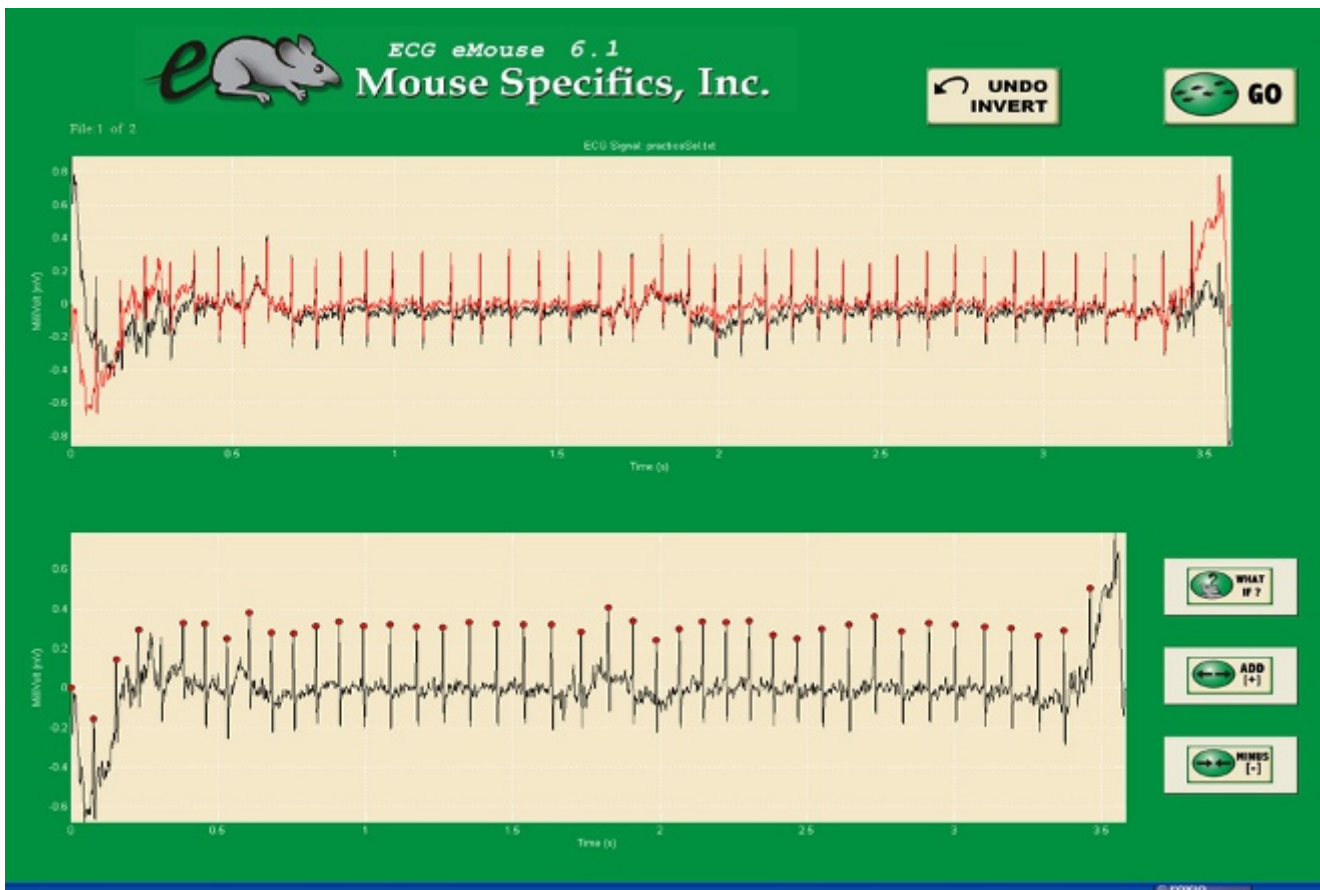


Figure C. Analysis phase, with the options to remove sections on the 'What if?' button below.



Parameters

	Version	Type	Req. Upload	Req. Analysis	Annotation	Increment	Option	Ontology Options	Derived	Unit	Data Type
Number of signals IMPC_ECG_001_001	1.2	simpleParameter	✓								FLOAT
HR IMPC_ECG_002_001	1.1	simpleParameter	✓		✓					bpm	FLOAT
CV IMPC_ECG_003_001	1.0	simpleParameter			✓					%	FLOAT
RR IMPC_ECG_004_001	1.2	simpleParameter	✓		✓					ms	FLOAT
PQ IMPC_ECG_005_001	1.0	simpleParameter			✓					ms	FLOAT
PR IMPC_ECG_006_001	1.1	simpleParameter	✓		✓					ms	FLOAT
QRS IMPC_ECG_007_001	1.2	simpleParameter	✓		✓					ms	FLOAT
ST IMPC_ECG_008_001	1.0	simpleParameter	✓		✓					ms	FLOAT
QTc IMPC_ECG_009_002	2.0	simpleParameter	✓		✓					ms	FLOAT
HRV IMPC_ECG_010_001	1.0	simpleParameter			✓					bpm	FLOAT
QTc Dispersion IMPC_ECG_011_001	1.0	simpleParameter			✓					ms	FLOAT
Mean SR amplitude IMPC_ECG_012_001	1.1	simpleParameter								mV	FLOAT
Mean R amplitude IMPC_ECG_013_001	1.1	simpleParameter								mV	FLOAT
rMSSD IMPC_ECG_014_001	1.0	simpleParameter			✓					ms	FLOAT
pNN5(6>ms) IMPC_ECG_015_001	1.2	simpleParameter								%	FLOAT
Waveform Image IMPC_ECG_025_001	1.0	seriesMediaParameter									IMAGE
Waveform Image Comment IMPC_ECG_026_001	1.0	simpleParameter									TEXT

Metadata

	Version	Type	Req. Upload	Req. Analysis	Annotation	Increment	Option	Ontology Options	Derived	Unit	Data Type
Equipment ID IMPC_ECG_016_001	1.0	procedureMetadata	✓								TEXT
Equipment Manufacturer IMPC_ECG_017_001	1.0	procedureMetadata	✓	✓			Mouse Specifics, Inc. AD Instruments World Precision Instruments				TEXT
Equipment Model IMPC_ECG_018_001	1.0	procedureMetadata	✓	✓			ECGenie ML870/p ML826/FE132 Iso-DAM8A ECGenie + gel pads ML866				TEXT
Anesthetic IMPC_ECG_019_001	1.0	procedureMetadata	✓	✓			Isoflurane Avertin Tribromoethanol No anesthesia				TEXT
Experimenter ID IMPC_ECG_020_001	1.0	procedureMetadata	✓								TEXT
Noise level IMPC_ECG_021_001	1.0	procedureMetadata									TEXT
Light level IMPC_ECG_022_001	1.0	procedureMetadata									TEXT
Date equipment last calibrated IMPC_ECG_023_001	1.1	procedureMetadata									DATE
Analysis Software IMPC_ECG_024_001	1.0	procedureMetadata		✓			eMouse Matlab				TEXT