Immunophenotyping IMPC_IMM_001

Purpose

This test differentiates immune cell sub-populations via flow cytometry.

Description: increased CD4-positive T cell number (MP:0008074), decreased CD4-positive T cell number (MP:0008075), etc..

Experimental Design

- **Minimum number of animals**: 3M + 3F
- **Age at test**: Week 16
- **Sex**: We would expect the results of this test to show sexual dimorphism

Equipment

Equipment

- Scissors and forceps for biopsy
- Precision balance
- Calibrated single and multichannel pipettes
- Plate shaker
- Refrigerated centrifuge
- Flow Cytometer (capable of distinguishing a minimum of 8 colours per well)
- Tissue dissociator:
  - GentleMACS tissue dissociator OR
  - Equipment for manual dissociation
- Cell counter equipment:
  - Orflo Moxi-Z Cell counter OR
  - Coulter Vicell XR OR Life Technologies Attune® Flow Cytometer

Supplies

- 96-well V-bottomed plates (Falcon #353263)
- Petri dishes
- Dispensing troughs
- Extra long 10 µl pipette tips for antibody solutions
- *(if using GentleMACS for dissociation)* C Tubes. It is acceptable to re-use these once.
- 50ml Falcon tubes
- Cell strainers e.g. 70m cell strainers that fit 50ml Falcon tubes (BD Falcon, #352350) OR Nytex
- Cell counter recipients (i.e., slides/cassettes/etc. for cell counter)
- *(if sample processing delayed)* RPMI 1640
• *(if sample processing on same day)* HBSS (with phenol red)
• CS (calf serum)
• PBS with Mg2+, with Ca2+ (for enzyme buffer used for DNAse and Collagenase D digestions)
• PBS without Mg2+, without Ca2+ (for FACS buffer to be used in all steps subsequent to enzymatic digest)
• EDTA (final concentration 2mM)
• Digestion enzyme (Collagenase D from Roche, #11088858001) stock solution in enzyme buffer (see below), aliquoted and stored at −20°C
• DNAses I stock solution (Sigma, #DN25) in enzyme buffer (see below), aliquoted and stored at -20°C
• RBC lysis buffer (eBioscience #00-4300-54 or BD Biosciences #555899, both 10X from manufacturer)
• HEPES (pH 7.2)

**Procedure**

This protocol requires several steps in the collection, preparation and analysis of the samples. Each one is detailed separately below.

**Reagent preparation**

*Note that two different PBS solutions are required for the protocol below, one with Ca2+ and with Mg2+, another without Ca2+ and without Mg2.*

- **Collection buffer:**
  - *(if spleens are to be processed on the same day)* HBSS with Ca2+/Mg2+ and phenol red (Life Technologies 14170161; check if it has phenol red) **OR**
  - *(if analysis will be delayed)* RPMI medium with 2% CS added.
- **FACS buffer** (for all steps subsequent to enzymatic digest; stable for up to 1 month in the fridge):
  - PBS 1X *without* Ca2+/Mg2+ **OR**
  - HBSS 1X *without* Ca2+/Mg2+
  - EDTA 2mM
  - 2% CS (v/v)
  - 10mM HEPES
- **Enzyme buffer** (for DNAse and Collagenase D digestions; Stable for up to 1 month in the fridge):
  - PBS *with* Ca2+ and Mg2+ **OR**
  - HBSS 1X *with* Ca2+/Mg2+
  - 2% CS (v/v);
  - 10mM HEPES
- **RBC Lysis buffer:** Prepare a 1X solution in ddH2O from lysis buffer.
- **Stopping buffer** (require 300 µl per sample):
  - 1x PBS without Ca2+ and without Mg2+ or HBSS
  - 0.1 M EDTA (37.5 g/L)
- **Antibody cocktails for Panels 1 & 2**
  - Protect antibodies and prepared cocktails from direct light.
  - Mastermix concentration, storage temperature and stability to be determined after panels 1 and 2 have been finalised and tested.
• Each sample will require 50 µl (or up to 100 µl) of diluted 1X antibody cocktail.
• Antibody cocktails should be gently but thoroughly mixed or quickly vortexed to
  ensure homogeneity of the solutions.
• In order to eliminate aggregated antibodies from your mix, centrifuge each
  antibody cocktail for 8 min at 20,000xg and 8°C prior to staining cells.

• **Read buffer / dead cell exclusion dye**
  • SytoxBlue at 1:10000 concentration in **FACS buffer** OR
  • SytoxGreen at 1:20000 concentration in **FACS buffer**
  • Zombie Near Infra-Red live dead from Biolegend at 1:2000 concentration
  • Require 200 l per well (i.e. 400 l for each spleen).

• **Enzyme cocktail (working solution):** 3 ml per each spleen, containing final
  concentrations of:
  • DNase I: 30 g
  • Collagenase D: 600 Mandl Units

  **NOTE:** To top up to the 3ml use enzyme buffer; any intermediate dilutions of the
  enzyme stock solutions should be prepared with enzyme buffer.

**Other preparations on the day**

• Bring RBC lysis buffer and stop solution to room temperature.
• Prepare wet ice box, label tubes, etc.

**Note** all centrifuge steps are: 5 min, 400 x g at 8°C

**Spleen collection**

• Collect the spleen from euthanized mice.
• Remove all fat from the spleen and weigh the organ on a petri dish (do not hydrate the
  organ before weighing it as this would lead to substantial errors in measurement).
• Place the spleen in a 1.5ml eppendorf tube with 1 mL of sample collection buffer on ice.
  Use:
  • *(if spleens are to be processed on the same day)* HBSS without calcium, without
    magnesium but with phenol red OR
  • *(if analysis will be delayed)* RPMI with 2% CS buffer.

**Spleen dissociation / digests**

  **If using a GentleMacs tissue dissociator:**

• Add the spleen to a GentleMACS C tube containing 3 ml of 1X enzyme cocktail.
• Clip the tube on GentleMACS dissociator and run programme spleen_2.
• Incubate cell suspension for 30 minutes with gentle mixing at least every 5 minutes.
  Register incubation temperature.
• Run programme spleen 3.
• Add 300 L of stopping buffer and mix by inversion to block enzymatic digestion and
  dissociate T cell-dendritic cell interactions.
• Filter cell suspension:
  • through 70 m Nylon mesh filter into a 50 mL Falcon tube OR
directly from C-tubes pour splenocyte suspension through 30 mm CellTrics Partec filters (#04-0042-2316) into 15 ml tubes.

(\textit{optional}) Wash the GentleMACS C tube with 5ml FACS buffer, filter and pool with flow-through from previous step.

- Centrifuge for 5 minutes, 400 x g at 8\(^\circ\)C and discard supernatant.

- Resuspend total splenocytes in 1 mL cold FACS buffer and keep on ice (this step is not required if counting is performed on the attune).

\textbf{OR, if performing manual digests:}

- Place weighed spleen in 12x75mm tube containing 1ml of collagenase solution in 1X HBSS with Ca\(^{2+}\) and Mg\(^{2+}\) (0.17-0.2 Wünsch unit/ml)
- Mince into fine pieces using small scissors, place on ice until all samples are minced.
- Add 2ml collagenase (0.17-0.2 Wünsch unit/ml) to each tube and place in a 37\(^\circ\)C water bath for 30 minutes.
- Tricturate (pipetting vigorously up and down using a 1 mL pipetman) the mixture to break up clumps.
- Spin at 500 x g in a swing bucket rotor for 5 min at 10\(^\circ\)C. Decant the supernatant, rack the tubes or vortex to resuspend the pellet. Add 2ml FACS buffer, mix well by vortexing, take 10 µl for the counting step.
- Dilutions for counting: 2 serial 1:10 dilutions (10µl cells + 90µl FACS buffer, then 10µl of the 1:10 dilution + 90µl buffer.)
- Spin for 5min, 500 x g at 10\(^\circ\)C, decant supernatant, blot the top of the tube, resuspend pellet at 1x10\(^8\) cells/ml.

\textbf{Cell counting}

- Perform a cell count on an aliquot of the re-suspended cells (adjust concentration according to the cell counter method used).
- Note down the cell count, correct for dilution and calculate the concentration in cells per µl.
- Cell count:
  - \textit{If performed before RBC lysis}, pipette the volume containing approximately 4 million cells/well to a 96 well plate in horizontal fashion starting from A1 onwards for panel 1 staining.
  - \textit{If performed after RBC lysis}, pipette the volume containing approximately 1-2 million cells/well to a 96 well plate in horizontal fashion starting from A1 onwards for panel 1 staining.
- Do the same for panel 2 staining in separate wells leaving a few empty rows between the panels to avoid cross contamination.
- Top up to final volume of 100 ml using FACS buffer, centrifuge, discard supernatant and keep plate on wet ice.

\textbf{Red blood cell lysis, blocking & staining}

- Remove plate from ice and add 30 to 100 ml of 1X RBC lysis buffer (at room temperature) to each cell pellet from the previous step.
- Pipette up and down 2-3 times to break up the pellet and ensure complete lysis. Alternatively, vortex the edges of the plates, then pipet quickly once to ensure resuspension is ideal for optimal lysis.
• Incubate for 1 minute at room temperature and then return to ice and add 100 to 200 ml of FACS buffer (to stop lysis) to each well.

**Note:** Following RBC lysis, every centrifugation step can be performed at 2000rpm for 1 minute in a 96 well plate, which significantly speeds up the protocol. Do take care to resuspend the cells very well to prevent HTS clumping.

• Centrifuge, discard supernatant and resuspend in 200 ml FACS buffer (this step is not required if lysis was performed in 30 µl, since there will be enough volume left in the well for a bigger wash of 200 µl; saves time on a spin).
• Again centrifuge and discard supernatant and resuspend in 50 ml of 1:100 Fc block and incubate on ice for 10 min. Top up to 200 ml using FACS buffer after incubation.
• Take antibody (AB) cocktails from the fridge. In order to eliminate aggregated ABs from your mix before use, centrifuge each AB cocktail for 8 min at 20,000 x g and 4°C.
• Centrifuge plate, discard supernatant and resuspend in 50 to 100 ml 1X AB mix in appropriate wells for individual panels followed by incubation on ice and in the dark for 20 min.
• **If using Sytox Blue/Sytox Green as live/dead discriminator:**
  • Top up to 200 ml with FACS buffer after incubation. Centrifuge, discard supernatant and resuspend in 200 ml FACS buffer.
  • When ready to read plate, centrifuge again and discard supernatant. Resuspend the pellet in 200 ml of read buffer (Sytox Blue diluted 1:10000 in FACS buffer; Sytox Green diluted 1:20000 in FACS buffer).
• **If using Zombie NIR dye as live/dead discriminator:**
  • Add 200 ml of PBS (RT) to all samples
  • Spin at 2000 rpm for 1 minute 8°C
  • Add 100 ml/well of Zombie Near-IR Live/Dead dye (1/2000) made up in PBS incubate at room temperature for 10 mins, add 200 ml FACS buffer.

**General Recommendations for Setting up Cytometer**

Set up the analyser to aim acquire 300,000 viable events (live cells) for each of Panels 1 and 2. 500,000 are recommended for panel 2 in order to increase robustness of myeloid population of low frequencies (macrophages, DCs).

**Gating Panel 1**

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<td>NKT cells (panel A)</td>
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<tr>
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<tr>
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## Parameters and Metadata

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- **Req. Upload:** true  
- **Is Annotated:** false
- **Unit Measured:** g

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**Percentage of live gated events in Panel A** IMPC_IMM_002_001 | v1.7

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- **Req. Upload:** false  
- **Is Annotated:** false
Unit Measured: %

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**T cells (panel A)** IMPC_IMM_003_001 | v1.5
simpleParameter


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**NKT cells (panel A)** IMPC_IMM_004_001 | v1.5
simpleParameter


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**NK cells (panel A)** IMPC_IMM_005_001 | v1.5
simpleParameter


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**Others** IMPC_IMM_006_001 | v1.4
simpleParameter

**CD4 T cells**  IMPC_IMM_007_001 | v1.4

simpleParameter


**CD8 T cells**  IMPC_IMM_008_001 | v1.4

simpleParameter


**DN T cells**  IMPC_IMM_009_001 | v1.5

simpleParameter


**DP T cells**  IMPC_IMM_010_001 | v1.2

simpleParameter

**CD4 NKT cells** IMPC_IMM_011_001 | v1.4

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**CD8 NKT cells** IMPC_IMM_012_001 | v1.5

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Is Annotated: false

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**DN NKT cells** IMPC_IMM_013_001 | v1.4

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**CD4 CD25+ T cells** IMPC_IMM_014_001 | v1.4

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Is Annotated: true

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**CD4 CD25- T cells** IMPC_IMM_015_001 | v1.4

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CD8 CD25- T cells  IMPC_IMM_017_001 | v1.4

DN CD25+ T cells  IMPC_IMM_018_001 | v1.5

DN CD25- T cells  IMPC_IMM_019_001 | v1.5
CD4 CD25+ NKT cells  IMPC_IMM_020_001  | v1.4
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CD4 CD25- NKT cells  IMPC_IMM_021_001  | v1.4
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CD8 CD25+ NKT cells  IMPC_IMM_022_001  | v1.5
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CD8 CD25- NKT cells  IMPC_IMM_023_001  | v1.5
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Is Annotated: true

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**CD4 CD44+CD62L+ T cells**  IMPC_IMM_029_001 | v1.2

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CD4 CD44-CD62L- T cells IMPC_IMM_031_001 | v1.2

CD8 CD44+CD62L- T cells IMPC_IMM_032_001 | v1.2

CD8 CD44+CD62L+ T cells IMPC_IMM_033_001 | v1.2

CD8 CD44-CD62L+ T cells IMPC_IMM_034_001 | v1.2
CD8 CD44-CD62L- T cells IMPC_IMM_035_001 | v1.2

simpleParameter


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DN CD44+CD62L- T cells IMPC_IMM_036_001 | v1.3

simpleParameter


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DN CD44+CD62L+ T cells IMPC_IMM_037_001 | v1.3

simpleParameter


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DN CD44-CD62L+ T cells IMPC_IMM_038_001 | v1.3

simpleParameter

**DN CD44-CD62L- T cells** IMPC_IMM_039_001 | v1.3

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**CD4 CD44+CD62L+ NKT cells** IMPC_IMM_041_001 | v1.2

**CD4 CD44-CD62L+ NKT cells** IMPC_IMM_042_001 | v1.3

**CD8 CD44+CD62L- NKT cells** IMPC_IMM_043_001 | v1.3

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**CD8 CD44+CD62L- NKT cells** IMPC_IMM_044_001 | v1.3
CD8 CD44+CD62L+ NKT cells IMPC_IMM_044_001 | v1.3

CD8 CD44-CD62L+ NKT cells IMPC_IMM_045_001 | v1.3

DN CD44+CD62L- NKT cells IMPC_IMM_046_001 | v1.2

DN CD44+CD62L+ NKT cells IMPC_IMM_047_001 | v1.2
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IMPC_IMM_048_001 | v1.3

**Percentage of live gated events in Panel B**  
IMPC_IMM_049_001 | v1.5

**Neutrophils**  
IMPC_IMM_050_001 | v1.2

**Monocytes**  
IMPC_IMM_051_001 | v1.2
Eosinophils IMPC_IMM_052_001 | v1.2

simpleParameter


NK Cells (panel B) IMPC_IMM_053_001 | v1.3

simpleParameter


NK Subsets (Q1) IMPC_IMM_054_001 | v1.3

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NK Subsets (Q2) IMPC_IMM_055_001 | v1.2

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NK Subsets (Q3) IMPC_IMM_056_001 | v1.2
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NK Subsets (Q4) IMPC_IMM_057_001 | v1.2
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NKT Cells (panel B) IMPC_IMM_058_001 | v1.2
simpleParameter


NKT Subsets (Q1) IMPC_IMM_059_001 | v1.2
simpleParameter


NKT Subsets (Q3) IMPC_IMM_060_001 | v1.2
T Cells (panel B) IMPC_IMM_061_001 | v1.2

T Subset IMPC_IMM_062_001 | v1.2

B Cells IMPC_IMM_063_001 | v1.2

B1B Cells IMPC_IMM_064_001 | v1.2
**B2B Cells**  IMPC_IMM_065_001 | v1.2

simpleParameter

**Follicular B Cells**  IMPC_IMM_066_001 | v1.2

simpleParameter

**Follicular B Cells (CD21/35+)**  IMPC_IMM_067_001 | v1.1

simpleParameter

**Transitional B Cells**  IMPC_IMM_068_001 | v1.3

simpleParameter
Transitional B Cells (CD21/35 low)  IMPC_IMM_069_001 | v1.2


MZB  IMPC_IMM_070_001 | v1.2


MZB (CD21/35 high)  IMPC_IMM_071_001 | v1.1


cDCs  IMPC_IMM_072_001 | v1.2

cDCs CD11b Type  IMPC_IMM_073_001 | v1.2

pDCs  IMPC_IMM_074_001 | v1.2

RP Macrophage (F4/80+)  IMPC_IMM_075_001 | v1.1

RP Macrophage (CD19- CD11c-)  IMPC_IMM_076_001 | v1.1

Equipment name  IMPC_IMM_077_001 | v1.0
**Anesthesia** IMPC_IMM_081_001 | v1.0

Option: Injection narcosis with Sodium Pentobarbital (Somnopentyl), none, Injection narcosis with Ketamine (100mg/kg)/Xylazine (10mg/kg), Injection narcosis with Tribromoethanol (Avertin), Isoflurane,

**Cell digestion** IMPC_IMM_082_001 | v1.0

Option: GentleMACS, manual, manual with needles,

**Cell digestion agent** IMPC_IMM_083_001 | v1.0

Option: Collagenase D, Collagenase II, Spleen Dissociation Kit,
Cell digestion agent manufacturer  IMPC_IMM_084_001 | v1.1

Options: Roche, Worthington, Gibco, Miltenyi Biotec, Sigma,

Cell digestion agent catalog number  IMPC_IMM_085_001 | v1.2

Options: #11088858001, CLS2LS004176, 17101-015, 130-095-926, C6885,

Cell counting  IMPC_IMM_086_001 | v1.1

Options: pre-lysis, post-lysis,

Cell counting equipment manufacturer  IMPC_IMM_087_001 | v1.0
Options: Life Technologies, American Optical, Beckman Coulter, BD Biosciences, Merck Millipore, Orflo, Nexcelom Bioscience, IntelliCyt,

Cell counting equipment model IMPC_IMM_088_001 | v1.0


Options: Countess Automated Cell Counter, Reichert Brightline, Gallios, BD LSR-II, Scepter, Attune, Moxi Z, 4468770, Cellometer Auto T4, iQue Screener PLUS,

Cell counting equipment name IMPC_IMM_089_001 | v1.0


Cell lysis buffer manufacturer IMPC_IMM_090_001 | v1.2


Options: eBioscience, BD PharmLyse, Jax, JMC, LONZA, home brew,
**Cell lysis buffer catalog number** IMPC_IMM_091_001 | v1.2

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**Options:** 00-4300-54, 555899, home brew, 10-548E,

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**Date and time of sacrifice** IMPC_IMM_092_001 | v1.0

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**Date and time of sample preparation** IMPC_IMM_093_001 | v1.0

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**Sample storage temperature until analysis (in Celsius)** IMPC_IMM_094_001 | v1.0

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</table>
Unit Measured: C

FCS repository reference (URL/ID) IMPC_IMM_095_001 | v1.0

Total number of acquired events in Panel A IMPC_IMM_026_001 | v1.4

Total number of acquired events in Panel B IMPC_IMM_027_001 | v1.2

Balanced salt solution type IMPC_IMM_096_001 | v1.0
**Balanced salt solution manufacturer** IMPC_IMM_097_001 | v1.0

*procedureMetadata*

**Options:** HBSS, PBS,

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**Balanced salt solution catalog number** IMPC_IMM_098_001 | v1.1

*procedureMetadata*

**Options:** Sigma, Life Technologies, Wisent, Wako, Gibco, Biochrom, home brew,

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**RPMI manufacturer** IMPC_IMM_099_001 | v1.0

*procedureMetadata*

**Options:** Sigma, Life Technologies, Jax, Wako, Gibco, none used,
**RPMI catalog number**  IMPC_IMM_100_001  |  v1.1

**Req. Analysis:** false  |  **Req. Upload:** true  |  **Is Annotated:** false

**Options:** R8758, 11875-101, home brew, 189-02145, 31800-022, none used, 11875-093,

**DNAse I manufacturer**  IMPC_IMM_101_001  |  v1.1

**Req. Analysis:** false  |  **Req. Upload:** true  |  **Is Annotated:** false

**Options:** Sigma, Spleen Dissociation Kit,

**DNAse I catalog number**  IMPC_IMM_102_001  |  v1.2

**Req. Analysis:** false  |  **Req. Upload:** true  |  **Is Annotated:** false

**Options:** DN25, D8764, 130-095-926,
Dead cell exclusion dye  IMPC_IMM_103_001 | v1.0

**Options:** Sytox Blue, Sytox Green, Zombie NIR, DAPI, Propidium Iodide,

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Dead cell exclusion dye manufacturer  IMPC_IMM_104_001 | v1.0

**Options:** Life Technologies, Biolegend, Sigma, home brew,

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Dead cell exclusion dye catalog number  IMPC_IMM_105_001 | v1.1

**Options:** S34857, S-34860, 423106, D9542, S11348, home brew, R37606, P4170,

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Cell digestion temperature (in Celsius)  IMPC_IMM_106_001 | v1.1
Panel A FCS file(s)  IMPC_IMM_107_001 | v1.0
seriesMediaParameter

Panel B FCS file(s)  IMPC_IMM_108_001 | v1.0
seriesMediaParameter

Automated analysis  IMPC_IMM_109_001 | v1.0
procedureMetadata
**Collection buffer manufacturer** IMPC_IMM_110_001 | v1.2

**Options:** Life Technologies,

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**Collection buffer catalog number number** IMPC_IMM_111_001 | v1.2

**Options:** 24020,

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**FACS buffer manufacturer** IMPC_IMM_112_001 | v1.1

**Options:** Life Technologies,
FACS buffer catalog number IMPC_IMM_113_001 | v1.1


Options: 14175,

Enzyme buffer manufacturer IMPC_IMM_114_001 | v1.1


Options: Life Technologies,

Enzyme buffer catalog number IMPC_IMM_115_001 | v1.1


Options: 14025,